Dynamic changes to signal allocation rules in response to variable social environments in house mice

Caitlin H Miller^{*}, Matthew F Hillock, Jay Yang, Brandon Carlson-Clarke, Klaudio Haxhillari, Annie Y Lee, Melissa R Warden, Michael J Sheehan^{*}

Department of Neurobiology and Behavior, Cornell University, Ithaca, NY, USA

*Authors for Correspondence:

Caitlin H Miller: chm79@cornell.edu Michael J Sheehan: msheehan@cornell.edu

Abstract

Male house mice use metabolically costly urine marks in intrasexual competition and mate attraction. Given the high costs of signaling and the depletable nature of urine reserves, males should dynamically modulate signal allocation as the social landscape is updated with new information. We investigate which aspects of male urine marking behavior are static or dynamic in light of changing social environments. To do this, we use thermal imaging to capture spatiotemporal data of urine deposition decisions. This novel method reveals fine-scale variation in urinary motor patterns in response to competition and social odors. Males demonstrate striking winner-loser effects in both the total allocation effort and temporal dynamics of scent marking. We find that competitive experience primes key temporal features of signal allocation and modulates responses to familiar and unfamiliar male scents. Males adjust their signaling effort, mark latency, and scent mark rhythm, depending on the scent identities present in the environment. Winners dramatically increase marking effort toward unfamiliar compared to familiar male scent, consistent with a 'dear enemy' effect. Losers, in contrast, greatly reduce marking to unfamiliar scent but increase marking effort to the scent of their familiar rival, consistent with a 'nasty neighbor' effect. Counter to the high lability of many features, the initial signal investment pattern influences allocation decisions days later, revealing the possibility of alternative scent mark strategies among competitive males. Thus, different features of urine mark signal allocation vary in responsiveness to fluctuating social landscapes, suggesting there are multiple distinct behavioral modules underlying marking behavior.

Keywords: scent mark, signal allocation, competition, winner-loser effects, dear enemy, nasty neighbor, familiarity, territory, decision-making, thermal recording

Introduction

1 2

3 Animals adjust their signaling behavior in response to recent experience and social context. Signalers may change 4 not only the frequency of signaling behavior, but also when, where, and how they signal in response to changing 5 social and physical environments (Hobson, 2020; Pasch et al., 2017; Patricelli & Hebets, 2016; Rauber & Manser, 6 2018; Sethi et al., 2019; Sullivan-Beckers & Hebets, 2014). Changes in allocation effort may allow individuals to 7 take advantage of signaling opportunities or avoid unprofitable signal investment (Alberts, 1992; Ferkin, 2015; 8 Gosling, 1982; Hurst & Beynon, 2004; Johnstone, 1996). In house mice (Mus musculus domesticus), males use 9 metabolically costly urine marks to mediate intrasexual competition and mate attraction. The abundance, spatial 10 distribution, and chemical composition of urine marks contain information about a male's competitive status and 11 identity (Desjardins et al., 1973; Drickamer, 2001; Ferkin, 2019; Gosling et al., 2000; Hurst, 1990; Hurst et al., 2001, 12 2005; Kaur et al., 2014; Lee et al., 2017; Nelson et al., 2015; Nevison et al., 2000). Previous studies have shown 13 that the total level of urine deposition is modulated by social dominance (Desjardins et al., 1973; Drickamer, 2001; 14 Hurst et al., 2005) and the presence of social odors in the environment (Hurst, 1990; Hurst et al., 2001; Kaur et al., 15 2014; Nevison et al., 2000). While urine marks convey rich social information, they are also directly depletable. Just as a car runs out of fuel, animals have a limited supply of urine to allocate toward signaling at any given moment. 16 17 Thus, urine marks pose a unique set of production constraints that are not observed in other well-studied signaling 18 systems, such as songs or visual displays (Cooper & Goller, 2004; Gil & Gahr, 2002; Laidre & Johnstone, 2013; 19 Ruppé et al., 2015). We explore the flexibility of signal allocation decisions, both on a moment-to-moment timescale 20 as well as over the course of days. Understanding how individuals integrate social information to allocate 21 depletable and costly signals has the potential to reveal fundamental features of complex decision-making 22 processes.

23 Male social relationships are shaped by competition and familiarity with conspecifics in house mice (Anderson 24 & Hill, 1965; Crowcroft & Rowe, 1963; Desjardins et al., 1973; Harrington, 1976; Koolhaas et al., 2013; Mackintosh, 25 1970: Poole & Morgan, 1976: Wolff, 1985). Urine marking mediates some of these relationships by allowing 26 assessment and recognition of individuals (Drickamer, 2001; Hurst et al., 2001, 2005; Kaur et al., 2014; Nevison et 27 al., 2000). Both stimulus familiarity and aggressive contests independently have strong effects on male urine 28 marking (Arakawa, Arakawa, et al., 2008; Arakawa, Blanchard, et al., 2008; Desjardins et al., 1973; Drickamer, 29 2001; Jones & Nowell, 1973), however it remains poorly understood how the two interact. One possibility is male 30 mice respond indiscriminately to the urine of other males (i.e. non-self) regardless of their experience with said 31 individuals. Another possibility is males respond differentially to the urine of males with whom they have established 32 relationships versus unfamiliar novel males. In many territorial species, familiar neighbors reduce aggressive 33 behaviors and signaling effort toward familiar individuals known as the "dear enemy" effect (Booksmythe et al., 34 2010; Briefer et al., 2008; Christensen & Radford, 2018; Tumulty & Bee, 2021; Zorzal et al., 2021). The dear enemy 35 effect is well-documented across vertebrate species, and is thought to lessen the costs of territorial defense 36 (Tumulty, 2018). Alternatively, marking may be influenced by a "nasty neighbor" effect (Christensen & Radford, 37 2018; Goll et al., 2017; Jin et al., 2021; Müller & Manser, 2007), where territorial males increase their signaling 38 effort toward familiar neighbors. Given the high costs and depletable nature of urine marks, males should 39 dynamically modulate signal allocation as the landscape is updated with new social information. The present study 40 aims to shed some light on these decision rules by exploring how established competitive relationships and identity 41 information influence male signal allocation decisions across social and scent-marked environments.

42 We investigate how males shift their signal allocation in response to an aggressive contest, to the presence of a 43 familiar male competitor, and to the presence of urine scent-marks of differing male identities. The objectives of this 44 study were to: (1) establish thermal imaging as a novel method for measuring scent marking, (2) explore how 45 competitive experience alters marking behavior, and (3) test the hypothesis that familiarity is important for signal 46 allocation decisions. To do this, we developed a 4-day trial design in which 31 pairs of age and weight-matched 47 breeding males of distinct wild-derived strains were paired as competitors and presented a series of social and 48 scent-marked trials (Figure 1A). Two wild-derived partially inbred house mouse strains were used to examine scent 49 marking behaviors, and ensured that males of each pair smelled distinct from one another (Kaur et al., 2014; 50 Sheehan et al., 2016, 2019). On the first day, paired males were placed in an arena separated by a mesh barrier (Figure 1A). Paired males could see, hear, and smell each other but were limited to minimal physical contact 51 through the mesh. The mesh barrier was subsequently removed, and males engaged in an aggressive contest or 52 53 "fight trial" (Figure 1A). Based on the total aggressive behaviors performed by each male in the fight trial, males 54 were easily and unambiguously classified as winners or losers (Figure S1). On the second day, each male was 55 placed in an empty open field arena without any stimuli aside from the arena itself (Figure 1A). On the third day, 56 males were placed back into the mesh arena with the same male competitor encountered on the first day (Figure 57 1A). Finally, on the fourth day, each male was exposed to one of four urine mark treatments. Each treatment 58 contained aliquoted male urine of three possible identities (self, familiar male, or unfamiliar male) in two spatially 59 distinct scent-marked zones (Figure 1A). The four treatment types span a range of scent mark combinations (self-





Figure 1. Experimental design and recording methods. (A) Trial design for a given pair of mice. All trials were 30 minutes long. Day 1: age and weightmatched males of distinct wild-derived strains were paired as competitors and placed into an arena and separated by a mesh barrier (dashed line). The mesh barrier was removed and males entered into an aggressive contest (fight trial) concluding in winning or losing males. Day 2: each male was placed into an empty open field arena. Day 3: males were placed back into the mesh arena with the same (familiar) male competitor from the first trial day. Day 4: each male was exposed to one of 4 possible treatments of aliquoted male urine of 3 possible identities (self, familiar and unfamiliar) into two urinemarked zones. The 4 treatment groups: self-self, self-familiar male, self-unfamiliar male, familiar male-unfamiliar male. The familiar male stimulus is the urine of a male's paired competitor (present in mesh and fight trials). (B) Thermal recording snapshot of a mesh trial (Days 1 & 3) depicting the regions of interest (ROIs: Wall vs. Barrier) used to score urine mark deposition and track space use. The dashed line indicates the mesh barrier separating the two males. The solid lines depict the ROIs each male can traverse on their side of the barrier. Urine marks are hot (orange-pink: close to the body temperature) on a cool (dark blue) ambient substrate (filter paper) temperature. The first and last minute of each scent-marking trial was trimmed prior to analysis (total analyzed trial length: 28 minutes) to minimize detection of startle-based urination events caused by placement of mice into arenas and any jostling caused during trial set-up and take-down. (C) Thermal snapshot of open field (Day 2) and urine-marked (Day 4) trials with the ROIs used for scoring (Corners vs. Center) indicated with solid lines. An example track of the mouse's trajectory two seconds before and after its current location is shown (light turquoise) with the mouse body's centroid indicated with a circle. Recently deposited urine marks (hot pink) align with the past track of the focal mouse. (D-E) Example final urine blot (on filter paper) of an open field trial imaged under UV light with urine marks fluorescing brightly (D), and the processed inverted urine blot image with urine marks shown in black (E). (F) Density plot depicting the temporal distribution of all thermally detected urine marks across all trials. (G) The total number of urine marks detected across trials using novel thermal imaging and standard UV blot imaging recording methods. A linear mixed model (LMM) was used to model the relationship between recording method and total urine marks detected. An analysis of variance (ANOVA) was used to test for the overall effect of recording method (significance code: NS p>0.05).

64 65 **Results**

65 66 67

Thermal imaging reveals the spatiotemporal dynamics of scent marking in real time

68 To best study urine allocation decisions in mice, we need to measure real-time spatial and temporal patterns of 69 scent mark deposition events. Mouse urine marking has been previously studied by capturing final snapshots of 70 marking patterns at the end of a trial, providing a cumulative output of behavior (Desjardins et al., 1973; Kaur et al., 71 2014). More recently, methods have been developed for tracking temporal changes in urine output by filming under 72 UV light or a moving paper floor (Hou et al., 2016; Keller et al., 2018). Here, we used thermal imaging as an 73 unobtrusive method for capturing in real time both the spatial and temporal allocation of urine marks by male house 74 mice (Figure 1). Urine leaves the body hot (close to body temperature) and quickly cools to below the ambient 75 substrate temperature, providing a distinctive thermal signature. In this study we examine both the where and the 76 when of urine mark deposition. Trials were performed on filter paper to present urine stimuli and to generate informative summary images for each trial by imaging the urine blots under UV light (Figure 1D-E). This further 77 allows for comparison of thermal recording with a traditional urine detection method. 78

79 Using thermal imaging we recorded a total of 9,314 urine deposition events across trials, and we explored the 80 temporal distribution of these depositions. We observe an initial spike in urine deposition with a peak of activity at 81 ~100s, followed by an exponential decline for the remainder of the trial length (Figure 1F). The majority (77%) of 82 marks are deposited within the first 15 minutes (800s) of the trial, suggesting males rapidly scent mark upon entering an environment (Figure 1F). Thermal imaging focuses on urine deposition, as marks are scored by the 83 distinct thermal profile of urine as it is deposited. UV light imaging cannot distinguish between deposition and 84 85 distribution events, as urine is further distributed by males walking through their own marks and tracking urine with 86 their paws and tail. However, UV imaging can also undercount urine marks when depositions occur in the same 87 location. UV light imaging at times detects more spots of urine than the true number of marks detected by thermal

90

91

imaging across trials, though not significantly ($F_{1,430} = 0.0034$, p = 0.95; Figure 1G). The two detection methods are also highly correlated (Figure S2), justifying the use of thermal imaging to examine how temporal urine allocation varies across contexts.



Figure 2. Male urine mark allocation in response to social competition across mesh trials. (A) Total urine marks deposited in Mesh 1 (pre-fight) and Mesh 2 (post-fight) by losers and winners. (B) Mesh trial urine blots of three paired male competitors (winner and loser) pre- and post-fight. (C) Linear mixed model (LMM) prediction of the total number of Mesh 2 marks (log-transformed) given fight outcome (winner=red; loser=blue) and initial signal investment (total number of Mesh 1 marks). (D) Histograms (top) of the temporal distribution of urine marks deposited by winners (red) and losers (blue) in Mesh 1 (pre-fight) and Mesh 2 (post-fight). Density plots (bottom) depict the density of urine mark deposition events over the 30-minute Mesh trials, distinguished by fight outcome (winner=red; loser=blue). (E) Linear mixed model (LMM) prediction of the latency to mark in both Mesh trials given the fight outcome (winner=red; loser=blue) and initial signal investment (total number of Mesh 1 marks). (A,C,E) Linear mixed models (LMMs) were used to model relationships, and analyses of variance (ANOVAs) were used to test for overall effects (significance codes: NS p>0.05, * p<0.05; ** p<0.01, *** p<0.01). Dependent variables were logarithmically transformed to meet assumptions for model residuals.

92

Competitive experience and initial signal investment shape urine mark allocation 93

94 Competitive social encounters can have a range of important consequences on the behavior and physiology of individuals (Harrison et al., 2018; Hsu & Wolf, 1999; Li et al., 2014; Milewski et al., 2022; Rose et al., 2017; Thomas 95 96 et al., 2015; Tibbetts & Crocker, 2014). How individuals respond to contest outcomes is often dependent on their assessment of their own resource holding potential, which is often further updated through experience (Arnott & 97 98 Elwood, 2009; Briffa & Elwood, 2009; Enguist & Leimar, 1983; Humphries et al., 2006), Signals play a key role in such encounters as they can convey information about the competitive ability of individuals (Kodric-Brown & Brown, 99 1984; Ligon & McGraw, 2016; Számadó, 2017; Tibbetts & Izzo, 2010). In house mice, initial scent mark investment 100 by males has been shown to contain information about their competitive ability (Drickamer, 2001). We therefore 101 102 predicted that (1) higher-marking males would be more likely to win aggressive contests, (2) winners would 103 increase while losers would decrease signaling after a contest, and (3) the changes in signaling after a contest may 104 depend upon their initial signaling strategies. We thus compared how males allocate urine marks in the presence of 105 a competitor before and after a fight by comparing the mesh trials on days 1 and 3 of the trial series (Figure 1A). 106 We classified winners and losers based on the total number of aggressive behaviors performed by each male in the 107 fight trial (Figure S1). In all contest pairings the fight outcome was overwhelming clear (Figure S1A), with winning 108 males performing significantly more aggressive behaviors than losing males ($t_{1,31}$ = -12.6, p = 1.09e-13; Figure 109 S1B). Fight outcome has a strong effect on total urine marks ($F_{1.68} = 10$, p = 0.0021), and there is a significant 110 interaction between fight outcome and trial number ($F_{1,60} = 12$, p = 0.0011). Before the fight (Mesh 1), the to-be winners include more high-marking individuals than the to-be losers, however, the two groups did not differ 111 112 significantly ($t_{1,112} = -0.69$, p = 0.88; Figure 2A). Initial signal investment may carry some information about 113 competitive ability (Drickamer, 2001), but this did not predict fight outcome between age and weight-matched males 114 in our trials. Post-fight (Mesh 2), the total urine marks deposited by winners is significantly higher than losers $(t_{1,112})$ 115 = -4.6, p = 0.0001; Figure 2A). Similar to previous studies (Arakawa, Arakawa, et al., 2008; Arakawa, Blanchard, et 116 al., 2008; Desjardins et al., 1973; Drickamer, 2001), this relationship appears to be largely driven by a decrease in 117 urine marking among losing males ($t_{1.61}$ = 3.3, p = 0.0059; Figure 2A,B).

118 We next assessed the role of initial signal investment (# Mesh 1 marks) and fight outcome on subsequent 119 allocation patterns (Figure 2C). Given prior research, we expected some males would mark highly, lose the fight, 120 and then suppress their marking behavior in the later trial (Desiardins et al., 1973). Instead, we found that the 121 importance of initial signal investment is more generalizable than this anticipated loser effect (Figure 2C). How 122 much an individual marked in the first mesh trial has a strong effect on the urine mark allocation in the second 123 mesh trial ($F_{1.59}$ = 9.2, p = 0.0036; Figure 2C). In other words, if you start off a low-marking individual you remain 124 relatively low-marking, regardless of the fight outcome. Accordingly, both high-marking losers and low-marking 125 winners are observed (Pair 3: Figure 2B). The pronounced winner-loser effects of urine mark allocation are 126 therefore strongly modulated by initial signal investment.

Losing has a notable effect not only the on number of marks, but also *where* individuals place those marks in the arena (Figure S3). On the second mesh trial, losers allocate their marks differently in space (at the wall vs. barrier) depending on whether they started off as high or low-marking (Figure S3A,B), suggesting that losers may be altering signaling strategies in addition to total signaling effort. Space use, on the other hand, does not differ (Figure S3C). All individuals spend more time in the social region of the arena (barrier) regardless of fight outcome (Figure S3C). Surprisingly, where males spend time does not correlate with where they mark (Figure S3D), indicating males are not simply marking where they spend time but are specifically allocating their urine marks.

135 Social experience influences the temporal dynamics of scent mark allocation

134

In addition to the total number of urine marks, mice may alter the relative timing of urine mark deposition, such that
marks may be more spaced out or clustered in time. The relative timing of urine deposition provides novel
information on the instantaneous rates of signaling, revealing how mice choose to spend their urine reserves. A
slow and regular mark deposition strategy is distinct from marking in rapid bursts upon the detection of a stimulus
or entry into an environment. In more natural contexts it is very plausible that rapid bursts of marking facilitate
competitive signaling, as males are traversing larger distances and navigating more complex social environments.

We inspected the distribution of urine mark deposition for winners and losers across mesh trials (Figure 2D). In the first trial, winners and losers display an initial peak in urine deposition at ~100 seconds (Figure 2D). Notably, the losers-to-be have a distinct second peak much later in the trial (~1000 seconds; Figure 2D). In the second mesh trial the effect of fight outcome is clear, with winners marking considerably more and losers less (Figure 2D). The density curves, however, reveal that both winners and losers allocate their marks earlier in the trial followed by a sustained decline in deposition events (Figure 2D). This shift in urine deposition regardless of fight outcome suggests a general priming effect of social competition on the temporal allocation of urine marks.

149 The timing of a male's first scent mark changes with fight outcome (Figure 2E). Mark latency is strongly influenced by initial signal investment ($F_{1,59}$ = 39, p = 4.4e-08). Similarly, trial order has a clear effect on the latency 150 151 to mark ($F_{1,57}$ = 10, p = 0.0021), while fight outcome does not ($F_{1,57}$ = 0.24, p = 0.62). The three-way interaction 152 between trial order, fight outcome, and initial mark investment significantly effects mark latency ($F_{1.58}$ = 10, p = 153 0.0021; Figure 2E). For both winners and losers, low-marking males are slower to mark than high-marking males, 154 characterizing a low and slow marking pattern on the first day. Conversely, high-marking individuals typically mark 155 rapidly upon entering the arena on the first day, representing a high and fast marking pattern. Across the two trial days, winners speed up their mark latency, though this effect is scaled to their initial mark investment (Figure 2E). 156 Initially high-marking losers have an increased latency after losing (i.e. they are slower to mark), but individuals who 157 158 marked infrequently on the first trial decrease their latency, demonstrating complex shifts in signaling behavior dependent on initial state. (Figure 2E). 159



Figure 3. Temporal dynamics of urine mark allocation across mesh trials. (**A**) Example data event plots depicting the urine marking of two pairs of male competitors over the course of both mesh trials for the entire trial duration (top) and a zoomed-in view of the first 200s (bottom). (**B**) Histograms of the intermark intervals (IMIs) for winners and losers in both mesh trials. Median values are indicated with dashed lines. The range of IMIs extends to nearly the full trial length (only the first 12s is shown). The maximum values are reported in the top right corner. Mesh 1: 65% of all IMIs are shown (< 12s), 57% of loser IMIs and 69% of winner IMIs. Mesh 2: 68% of all IMIs are shown (< 12s), 51% of loser IMIs and 72% of winner IMIs. (**C**) Box and violin plots of within-bout IMIs by fight outcome and mesh trial. (**D**) Donut plots by trial and fight outcome depicting the proportion of bouts composed of: 1 mark, 2 marks or 3+ marks. (LMMs) were used to model relationships, analyses of variance (ANOVAs) were used to test for overall effects, and post hoc pairwise comparisons were performed using the *emmeans* package (significance codes: NS p>0.05, * p<0.05; ** p<0.01, *** p<0.001).

We next examined the timing and rhythm of urine marking across entire mesh trials. Event plots depicting the timing and frequency of marks deposited for two male pairs are shown (Figure 3A). The winning males increase the front-loading of urine deposition in the second trial, whereas the losers decrease overall mark deposition but still mark relatively quickly in the second trial (Figure 3A). Examining the timing of marks across the trial reveals unanticipated patterns as well. The intervals between marks differ noticeably between the first and second mesh

165 trials, particularly when marks are made in close sequence to each other. In the first trial, the mark sequences have 166 larger intervals, producing chains of marking events (Figure 3A & S4). Whereas in the second mesh trial the mark 167 sequences are compressed into shorter intervals, creating bursts of marking events (Figure 3A). To examine this relationship further, we inspected the distribution of inter-mark intervals (IMIs) among winners and losers for both 168 169 trials (Figure 3B). On the first trial the median IMI of winners and losers is similar (Figure 3B). However, on the 170 second trial, the median IMI for losers increases, driven by the fact that fewer marks are being made overall. While 171 IMIs have a wide range, the most frequent mark intervals are under 3 seconds (Figure 3B & S4). In the first trial, the 172 most frequent IMIs are less than 3 seconds for winners and losers, though winners have a lower median mark 173 value. (Figure 3B). In the second trial, there is a clear peak of IMIs of less than 1 second for both winners and 174 losers (Figure 3B). This suggests that both winners and losers are producing mark "chains" on the first trial and 175 "bursts" of marks on the second. The overall median IMI interval is unchanged for winners but increases notably in 176 losers.

To explore this shift in temporal dynamics *within* urine mark sequences, we classified marks that occur in intermark intervals of less than 3 seconds as marking bouts (Figure S4). Bouts can thus consist of a single mark or a series of marks. We then examined the variation in IMIs within urine mark bouts (Figure 3C). As expected, trial has a strong effect on within-bout IMIs (i.e. IMIs for bouts with 2+ marks, $F_{1,428}$ =304, p = 2.0e-16) while fight outcome does not ($F_{1,46}$ = 0.079, p = 0.78; Figure 3C). Thus, marking events within bouts are more rapid in the second trial, regardless of whether they won or lost, indicating that competitive experience may prime a particular marking motor pattern, regardless of fight outcome.

184 We next inspected whether bouts are composed of 1 mark, 2 marks or 3+ marks (Figure 3D). On the first trial 185 losers have more single-mark bouts and winners have more multi-mark bouts (Figure 3D). This relationship 186 becomes even more stark in the second mesh trial, as losers decrease the overall number of marks across mesh 187 trials, but the bout composition remains very similar (Figure 3D). Winners, on the other hand, increase the number 188 of marks and alter their bout composition to include more multi-mark bouts (Figure 3D). To further explore bout 189 composition, we compared the average number of marks per bout by fight outcome and trial (Figure 3E). In 190 contrast to the IMIs of urine mark bouts, bout composition is strongly affected by fight outcome ($F_{1.58} = 10, p =$ 191 0.0022; Figure 3E). On the second trial, winners have a significantly higher average number of marks per bout than 192 losers ($t_{1,111} = -3.0$, p = 0.012; Figure 3E). This dataset reveals striking patterns of signaling behavior in male house 193 mice that would have otherwise gone unnoticed without temporal data possible from thermal imaging. 194



Figure 4. Urine mark allocation across scent-marked contexts in response. **(A)** Total urine marks deposited by winning and losing males in the open field trial (OFT) and the four urine-marked treatments: self-self (S-S), self-familiar male (S-FM), self-unfamiliar male (S-UM) and familiar male-unfamiliar male (FM-UM). All males experienced the OFT. Males experienced only one of the four urine-marked treatments. **(B)** Schematic of the urine stimulus components for the OFT and urine-marked treatments. OFTs are "no stimulus" trials (grey), S-S and S-FM have "no unfamiliar male" urine present (purple), and S-UM and FM-UM trials have "unfamiliar male" urine present (orange). **(C)** The difference in total marks deposited by males in the Marked trials relative to the empty OFTs (logarithmically transformed). Urine-marked treatments are grouped as "no unfamiliar male" urine (purple: S-S and S-FM) and "unfamiliar male" urine (orange: S-UM and FM-UM). Post hoc pairwise comparison significance values indicated at the top of boxplots. One-sample tests (deviation from 0) significance values are indicated on the bottom of the boxplots **(D)** Urine mark density plots of losing and winning males toward an empty arena (OFT), to trials with no unfamiliar male urine: S-S (light purple) and S-FM (dark purple), and to trials with unfamiliar male urine: S-UM (light orange) and FM-UM (dark orange). **(E)** Linear mixed model (LMM) prediction of the latency to mark in the OFT trials (gray), Marked trials with "No Unfamiliar" male urine (purple), and Marked trials with "No initial signal investment (total number of Mesh 1 marks). **(A, C, E)** Linear mixed models (LMMs) were used to model relationships, analyses of variance (ANOVAs) were used to test for overall effects, and post hoc pairwise comparisons were performed using the *emmeans* package (significance codes: NS p>0.05, * p<0.05; ** p<0.01, *** p<0.01). Dependent variables were logarithmically transformed to meet assumptions for model residuals.

195

196 Dominance and familiarity interact to shape countermarking behavioral dynamics

197 Given that males dynamically adjust marking behavior in response to social competition, we next explored 198 allocation decisions toward the scent marks of other males. We were especially interested in whether males use 199 knowledge of a recent competitor's identity in their signaling decisions, as males will competitively counter-mark to

200 (i.e. mark over) the urine marks of other males (Hurst & Beynon, 2004; Kaur et al., 2014). While it is well-201 established that males alter marking behavior in response to fight outcome (Desjardins et al., 1973; Drickamer, 202 2001; Hurst, 1990) and can finely discriminate urine identities (Hurst et al., 2001; Kaur et al., 2014), we have a 203 limited understanding of how males implement this information in a competitive marking context. Do males adjust 204 their scent marking behavior depending on their relationship to a male competitor? What role does familiarity play in 205 signal allocation dynamics? We hypothesized that fight outcome would shape urine marking behavior even in the 206 absence of a male competitor, and that male identity in urine-marked trials would strongly govern signal allocation 207 decisions.

208 To address these questions, we compared two trial types within the trial series in which no conspecifics were 209 present: open field trials (OFTs) and urine-marked trials (Figure 1A). The OFT contained no stimuli, while the urine-210 marked trials each contained two spatially distinct urine-marked zones of specific identities: their own urine (self: S), familiar male (FM) competitor urine, and/or unfamiliar male (UM) urine (Figure 4B). UM urine was collected from a 211 212 different strain (C57BL/6J) and pooled to ensure a distinct urine profile with no individual-specific effects on the UM 213 stimulus. We examined responses to an empty arena (OFT) and to the four different urine stimulus sets; S-S, S-214 FM, S-UM and FM-UM (Figure 1A) by fight outcome and initial signal investment. Trial type ($F_{4,76}$ = 5.2, p = 215 0.00089), fight outcome ($F_{1,83}$ = 27, p = 1.6e-06), and initial signal investment in the first trial ($F_{1,58}$ = 32, p = 4.2e-216 07), all significantly effect marking behavior of males (Figure 4A). As does the two-way interaction between trial type and fight outcome ($F_{4,77}$ = 5.8, p = 0.00041). Winners tend to mark more than losers, and losers typically mark 217 218 lowly across treatment types. This pattern is further observed in the empty arena trials ($t_{1,100} = -3.9$, p = 0.0034; 219 Figure 4A). Notably, winners and losers show opposite responses towards familiar versus unfamiliar urine. 220 Treatments with only familiar urine (Figure 4A,B, purple: S-S and S-FM) exhibit comparable marking responses in 221 winners and losers (Figure 4A). While it's perhaps less surprising that winners and losers mark comparably lowly to 222 their own urine (S-S; $t_{1,99}$ = -0.83, p = 1.00), it is striking that winners and losers do not differ in their response to the 223 S-FM treatment ($t_{1,105} = -0.44$, p = 1.00; Figure 4A). The opposite pattern is observed in the presence of unfamiliar 224 urine. Winners mark significantly more than losers to both S-UM ($t_{1.108}$ = -3.6, p = 0.0091) and FM-UM ($t_{1.109}$ = -6.0, 225 p < 0.0001) treatments (Figure 4A). In trials with two different urine identities, we had anticipated males would 226 differentially allocate urine towards each marked corner, though we did not detect such differences in either winners 227 or losers (Figure S5A). While scoring the trials, it became clear that the space was too small to delineate corner-228 based stimuli as distinct ROIs like we had originally planned. Especially since males frequently walk through 229 corners while performing a scent-mark bout that extends across multiple ROIs. Furthermore, our results indicate 230 males may respond to the most 'extreme' (or most salient) odor information in the environment within the spatial 231 confines provided (Figures 4 & 5). However, space use patterns of losing males suggest that mice are detecting 232 differences in the scent marks (Figure S5B). Losers spend less time in the center ROI compared to winners ($t_{1,230}$ = 233 -3.7, p = 0.0069), and losers spend less time in UM-marked corners relative to empty ones ($t_{1.199}$ = -3.5, p = 0.0012) 234 (Figures 1C & S5B).

235 We observed very similar responses in the two treatments with unfamiliar urine present (S-UM and FM-UM) as 236 well as the two treatments without unfamiliar urine (S-S and S-FM), so we collapsed these similar treatments 237 (purple: no unfamiliar male, orange: familiar male) to further explore the role of familiarity and fight outcome on 238 signal allocation (Figures 4B-E). We standardized the marking behavior of males by calculating the difference in 239 marks made in an empty arena (OFT) relative to a scent-marked environment (Figure 4C). The interaction between 240 fight outcome and familiarity strongly shapes marking behavior in scent-marked contexts ($F_{1.58} = 13$, p = 0.00054; 241 Figure 4C). Winners increase the number of marks significantly more than losers in trials with unfamiliar urine 242 present ($t_{1,58}$ = -3.0, p = 0.0069), whereas winners and losers do not differ when familiar-only scent marks are 243 present ($t_{1.58}$ = 2.0, p = 0.17; Figure 4C). What is also apparent is the inverse response of winners and losers 244 toward familiarity (Figure 4C). Winners mark relatively more in the presence of unfamiliar urine and lowly to familiar-245 only urine $(t_{1.58} = -3.0, p = 0.014)$, while losers mark lowly in the presence of unfamiliar urine and highly to familiar-246 only urine $(t_{1.58} = 2.2, p = 0.12;$ Figure 4C). Notably, losers in the familiar-only treatment $(t_{1.14} = 4.5, p = 0.00048)$ 247 and winners in the unfamiliar treatments ($t_{1,16} = 4.4$, p = 0.00041) deviate significantly from zero, while their 248 opposing treatments do not (Figure 4C).

249

250 Temporal variation in signal allocation during countermarking

251 The timing of signal allocation in scent-marked environments was examined (Figure 4D). We first looked at the 252 distribution of deposition events across trials (Figure 4D). In trials with no urine stimulus (OFTs), winners allocate 253 marks early in the trial (peak density ~100s), while losers mark less with a later peak at ~250s (Figure 4D). In 254 contrast, winners and losers have remarkably similar density curves for urine-marked trials containing familiar-only 255 urine (purple: S-S and S-FM) (Figure 4D). For S-S trials, both winning and losing male density curves display a 256 single initial peak (~100s), whereas the S-FM trials' density curves reveal an earlier initial peak (~70s) and a 257 smaller second peak later in the trial (Figure 4D). In trials with unfamiliar urine, winners and losers differ 258 dramatically. Winning males deposit large amounts of urine very quickly, creating a large initial spike in the density curves in both S-UM (light orange) and FM-UM (dark orange) treatments (Figure 4D). Losing males drop off and
 slow down their urine mark deposition considerably, generating density curves with small and delayed peaks
 (Figure 4D). The temporal distribution of urine marks is thus modulated by fight outcome and familiarity in scent marked environments.

As the temporal dynamics of scent-marks were overlapping in trials with or without unfamiliar male urine, we 263 264 collapsed these into treatment groups (Figure 4E). We further modeled the effects of treatment group, fight 265 outcome, and initial signal investment, on the latency to mark (Figure 4E). Mark latency is significantly predicted by 266 the number of marks made in the first mesh trial, i.e. the initial investment recorded 3 days earlier ($F_{1.57}$ = 10, p = 267 0.0024; Figure 4E). For winners and losers, initially low-marking individuals are slower to mark, and initially high-268 marking individuals are faster to mark (Figure 4E). This relationship is most stark among winners, which exhibit 269 steep slopes across treatment groups, while losers display more modest slopes (Figure 4E). The interaction 270 between fight outcome and initial signal investment, however, is moderate ($F_{1.57}$ = 3.7, p = 0.056; Figure 4E). The 271 effect of fight outcome on mark latency is not significant ($F_{1.56} = 1.3$, p = 0.25; Figure 4E). Treatment group on the 272 other hand, significantly effects the speed of marking response ($F_{1,75} = 3.2$, p = 0.048; Figure 4E). Losers mark 273 most rapidly in familiar-only trials, and winners mark most rapidly in trials with unfamiliar urine (Figure 4E). The 274 intersection points of the linear models for winners and losers reveal further insights. Winners transition to a more 275 rapid marking response relative to losers differently across treatments groups depending on initial signal 276 investment. In familiar-only trials, only the initially very high-marking (>85 marks) winners mark more quickly than losers, others are slower to mark. The opposite is true in trials with unfamiliar urine, in which even initially low-277 278 marking (>20 marks) winners mark more rapidly than losers (Figure 4E). This demonstrates that initial signal 279 investment has long-term power for predicting marking behavior, including the temporal allocation of urine marks. 280



Figure 5. Temporal dynamics of urine signal allocation across scent-marked contexts. **(A)** Example data event plots depicting urine marking in OFT and Marked trials of four losing males, each exposed to one of the four different scent-marked treatments: self-self (S-S), self-familiar male (S-FM), selfunfamiliar male (S-UM), and familiar male-unfamiliar male (FM-UM). The event plot for the entire trial duration is shown on top and a zoomed-in view of the first 200s is shown below. **(B,C,D)** Urine-marked treatments are grouped as "no unfamiliar male" urine (purple: S-S and S-FM) and "unfamiliar male" urine (orange: S-UM and FM-UM). **(B)** Box and violin plots of within-bout IMIs by fight outcome and trial group: OFT, No Unfamiliar (S-S & S-FM), and Unfamiliar (S-UM & FM-UM). **(C)** Donut plots by trial and trial group depicting the proportion of bouts composed of: 1 mark, 2 marks or 3+ marks. Mark totals in the bottom left-hand corner. **(D)** Example data event plots depicting urine marking in OFT and Marked trials of four winning males, each exposed to one of the four different scent-marked treatments: S-S, S-FM, S-UM, FM-UM. The event plot for the entire trial duration is shown on top and a zoomedin view of the first 200s is shown below. **(E)** Boxplot of the average number of marks per bout by fight outcome and trial group. **(B,E)** Linear mixed models (LMMs) were used to model relationships, analyses of variance (ANOVAs) were used to test for overall effects, and post hoc pairwise comparisons were performed using the *emmeans* package (significance codes: NS p>0.05, * p<0.05; ** p<0.01, *** p<0.01).

281 We next examined the timing and composition of marking bouts (Figure 5). More chain-like bouts are observed 282 in OFTs, whereas more rapid bursts of urine marking are produced in scent-marked trials (Figures 5A, 5D & S5C). 283 Therefore, over the 4-day trial series males mark increasingly in bursts, suggesting competitive experience shapes 284 temporal features of signal allocation. To explore this further we looked at within-bout IMIs (Figure 5B). Both fight 285 outcome ($F_{1.64} = 6.2$, p = 0.015) and treatment group ($F_{1.152} = 40$, p = 1.2e-14) significantly effect within-bout IMIs, 286 with a modest interaction ($F_{1,154} = 2.5$, p = 0.083; Figure 5B). As expected, the within-bout IMIs are significantly 287 longer in OFTs than either scent-marked treatment groups among winners or losers (Figure 5B). Winners, however, 288 marked with similar rapid bursts (short IMIs) regardless of familiarity with the urine stimulus (Figure 5B).

289 Conversely, losers tend to mark in bursts specifically during familiar-only trials (Figure 5B). This bout timing is most 290 prominent in the S-FM trials (Figure 5A & S5C), which reveals losers distinctly allocate their urine marks based on 291 the identity of urine marks in the environment. It is striking that, again, losers signal most conspicuously toward 292 males who recently defeated them in a competitive contest.

293 The number of marks per bout changes with social outcome and scent-mark type (Figure 5C). The average 294 number of marks deposited per bout is significantly shaped by scent-mark familiarity ($F_{1.76} = 13$, p = 1.3e-05) and 295 fight outcome ($F_{1,66} = 22$, p = 1.9e-05), with a strong two-way interaction ($F_{1,76} = 6.3$, p = 0.0031; Figure 5E). Fight 296 outcome and familiarity both influence the composition of marking bouts (Figures 5C & 5E). In an environment 297 empty of scent marks (OFT), winners allocate considerably more multi-mark bouts than losers (30% vs 5%; Figure 298 5C). And the average number of marks per bout is significantly higher among winners ($t_{1,105}$ = -3.0, p = 0.027; 299 Figure 5E). Interestingly, the differences in bout composition narrows in scent-marked trials with familiar-only urine 300 (Figure 5C). In these trials, winners deposit slightly more multi-mark bouts (38%), while losers dramatically shift the 301 amount of multi-mark bouts (26%; Figure 5C). The average number of marks per bout similarly does not differ 302 between winners and losers in familiar-only trials ($t_{1,117} = -1.0$, p = 0.87; Figure 5E). The reverse is true for trials with 303 unfamiliar male urine present (Figure 5C). Here, losers produce bouts with similar bout compositions to the empty 304 OFTs (Figure 5C). Winners double the proportion of multi-mark bouts compared to the OFTs (60%), and many of 305 these bouts contain at least 3 marks (46%; Figure 5C). Expectedly, the average number of marks per bout is 306 significantly higher among winners when unfamiliar urine is present ($t_{1,117}$ = -5.6, p = <0.0001; Figure 5E). Thus, the 307 dynamic temporal patterns of urine allocation change in response to the identity of urine to countermark.

309 **Discussion** 310

308

311 Taking advantage of a novel thermal imaging approach, we uncovered dramatic shifts not only in the amount of 312 urine marking but also the relative timing of urine deposition in response to varying social environments and recent social experiences. In contrast to the dynamic changes observed for some features of scent marking, we found that 313 314 initial marking strategy significantly explained allocation efforts days later. Our study reveals a mixture of static and 315 dynamic features of urine marking behavior in response to different social contexts. At the start of trials, all males 316 were fully adult and sexually mature with similar prior experiences. Nevertheless, we found substantial variation in 317 the amount, latency, and timing of scent marking in the first trial (Figures 2 & 3). Competitive social experiences 318 had multiple effects on scent marking behavior including priming effects on scent mark timing as well as winner and 319 loser effects. The temporal rhythms of scent marking behavior changed over the course of the trials and indicate a 320 dynamic and socially responsive feature of signaling behavior that had been previously unobserved.

321 Winning or losing an aggressive contest has strong and long-lasting effects on signaling decisions (Figures 4 & 322 5), consistent with classically described winner-loser effects (Dugatkin, 1997; Harrison et al., 2018; Hsu & Wolf, 323 1999). These winner-loser effects most prominently alter total allocation effort and marking bout composition. As 324 described in the literature, we find males quickly downregulate urine allocation after losing a competitive contest 325 (Arakawa, Blanchard, et al., 2008; Desjardins et al., 1973). Though signal allocation is influenced by recent social 326 outcomes, we find that initial signal investment has both stable and robust effects on marking behavior. In other 327 words, where males start off influences their signaling decisions days later. Low-marking individuals remain 328 relatively low-marking, and high-marking individuals stay relatively high-marking. The magnitude of the observed 329 winner-loser effects is therefore contingent on the initial investment decisions of males.

330 We detect "silent" low-marking winners in our dataset, for which we could find no previous description in the 331 literature. It's noteworthy that studies do sometimes pre-screen males for baseline urine marking behavior in scent-332 marking assays, which we did not do (Kaur et al., 2014). The detection of multiple apparently competitive males 333 that mark at very low rates suggests several possible hypotheses to explain their low marking behavior. First, the 334 result may be driven in part by our trial design. By pairing evenly-matched males, it could be we observed contests 335 in which low-marking males won a fight because two relatively low-marking individuals were paired together. 336 Additionally, better-than-expected outcomes could give rise to slower response times than is observed for worse-337 than-expected outcomes, in which high-marking losers rapidly downregulate signal allocation (Arakawa, Blanchard, 338 et al., 2008; Desjardins et al., 1973). Second, the low-marking males may differ in some aspect of hydration 339 physiology that we did not measure. Species and strains of mice vary in water intake and urination levels (Bittner et 340 al., 2021; Fertig & Edmonds, 1969; Moro & Bradshaw, 1999), though we observed low marking winner males from 341 both strains used in this study. Third, being a "silent" yet competitive male might represent a distinct signaling 342 strategy in house mice. Given the high metabolic costs of signaling, it's plausible that some males withhold signal 343 investment to continue investing in body mass or to avoid detection by other males. House mice males may pursue 344 diverse strategies, including the classically described "territorial male" that invests highly in urine marking and 345 territory as well as scent-silent "sneaker males" (Aubin-Horth & Dodson, 2004; Bhandiwad et al., 2017; Miles et al., 346 2007; Sinervo & Lively, 1996; Zamudio & Sinervo, 2000). While our data does not directly test this relationship, the 347 frequency of low-marking winners warrants further investigation. Certainly, the simple correlation between marking

and dominance is considerably more complex than previously described, and the interaction between competitive
 social experience and initial investment is crucial to understanding male signaling decisions.

350 Several features of marking behavior are primed by competitive experience. Mice mark more rapidly after a 351 contest, regardless of outcome (Figure 2). The time between deposition events similarly shrinks, such that marking 352 bouts transition from chain-line mark sequences to rapid bursts, resulting in fundamental shifts in motor pattern 353 sequences. Aggressive contests thus appear to push males into a competitive state, driving changes in fine motor 354 adjustments in marking behavior. Voluntary, involuntary, and context-dependent urination are all mediated by 355 neuronal subpopulations in Barrington's nucleus in the brainstem (Hou et al., 2016; Keller et al., 2018; Verstegen et 356 al., 2019). The fine-scale modulation of urinary motor control we observe in our dataset adds additional complexity 357 to this underlying circuitry, as our data reveal social interactions modulate the rhythms of urination behavior. This 358 result opens the possibility for future work to examine how social experience influences finely-controlled motor 359 outputs in an important social signal. Particularly as in more natural environments the rhythm and timing of urine 360 deposition may be crucial for efficient signal allocation as males traverse large distances and defend territories.

361 Males show markedly different responses to urine based on their familiarity with the scent and their recent fight 362 outcome. Past studies have shown that males can finely distinguish self from non-self urine (Kaur et al., 2014) and 363 that female mice recognize specific males based on their urine marks (Hurst et al., 2001, 2005). We had designed 364 trials with scent-marked corners with urine of different males expecting to see evidence that males allocated marks differently to each corner. We did not detect clear effects of differential allocation to marked corners, suggesting two 365 366 non-mutually exclusive possibilities: (1) the scent of unfamiliar males may be much more important in driving 367 allocation decisions and (2) the size of the arena may be too small for the marks to be in appreciably different 368 locations from the perspective of the mouse. Established competitive relationships, or familiarity, with another male 369 profoundly shapes marking behavior. Familiarity affects most facets of urine marking, including: total allocation, 370 timing of deposition, and marking bout composition. Surprisingly, losers tend to increase mark allocation effort and 371 display more frequent bursts of multi-mark bouts in response to familiar scent marks. In contrast, winners 372 downregulate their marking efforts toward familiar urine marks. Male mice are known to increase countermarking 373 toward non-self male urine (Hurst & Beynon, 2004; Kaur et al., 2014); however, we find that this effect is strongly modulated by recent experience (or lack thereof) with the urine's producer. We find evidence for the dear enemy 374 375 effect in winners and the nasty neighbor effect in losers, suggesting that recent social experiences modulate how 376 animals invest in territorial advertisement and signaling. The dear enemy effect has been reported in a wide array 377 of species (Booksmythe et al., 2010; Briefer et al., 2008; Christensen & Radford, 2018; Siracusa et al., 2021; 378 Tumulty & Bee, 2021; Zorzal et al., 2021). In our study, the responses toward familiar males are even more stark 379 when compared to how males respond to unfamiliar urine marks. Winners dramatically upregulate all competitive 380 marking efforts and losers essentially go completely scent "silent." Males remain aggressive and vigilant toward 381 unfamiliar or new neighboring males under the dear enemy model, and winners in our trials aggressively mark 382 toward novel males as well. Novel males would represent a threat to their current dominance status, motivating 383 winning males to invest further in signaling behaviors. Losers on the other hand are at risk of an additional 384 aggressive encounter after being recently defeated in a contest. By staying "silent" losers may avoid further conflict 385 with a new territorial contender, potentially in a "wait-and-see" strategy (Olivier et al., 1991; Rychlik & Zwolak, 386 2005). However, when presented with urine of the male that recently defeated them, losers actually upregulate 387 marking efforts. This response may be a "nasty neighbor" effect, in which the threat of familiar territorial males 388 exceeds that of strangers (Christensen & Radford, 2018). Alternatively, this increased marking response in losers 389 could be a form of subordinate marking to mediate recognition and thereby reduce aggression with a familiar 390 competitor. Male house mice thus actively track multiple identities in the environment and dynamically adjust their 391 signaling decisions in terms of total allocation and timing.

This work underlines the importance of examining signal responses across variable social odor landscapes and understanding the decision rules behind costly and complex behaviors. Because thermal recording allows for highthroughput and detailed analysis of scent marking, future studies can explore even more detailed social contexts and odor environments. This is particularly relevant given that our results provide new evidence for possible alternate signaling strategies, as well as complex territorial dynamics reflecting dear enemy and/or nasty neighbor effects in house mice. Lastly, the implementation of this novel thermal recording method has the potential to reveal important features underlying the neurophysiological basis of socially-modulated and voluntary urination behaviors.

399

400 Materials and methods

401

402 Key resources table

Reagent or resource	Source or reference	Identifiers		
Experimental Models: Organisms/Strains				
NY2 wild-derived inbred house mouse strain (<i>M. m. domesticus</i>)	Sheehan Lab, Cornell University (Ithaca, NY)	NA		

NY3 wild-derived inbred house mouse strain (<i>M. m. domesticus</i>)	Sheehan Lab, Cornell University (Ithaca, NY)	NA			
C57BL/6J (<i>M. m. domesticus</i>) The Jackson Laboratory		JAX: 000664			
Recording Systems					
PI 640 Optris infrared cameras	Optris Infrared Sensing (Portsmouth, NH)	https://www.optris.com/infrared-cameras			
iDVR-PRO CMS IR security cameras	CCTV Camera Pros	https://www.cctvcamerapros.com/DVR- Remote-Access-non-IE-s/470.htm			
Materials & Solutions					
Whatman filter paper (46 x 57 cm)	Sigma-Aldrich	1002-917			
Colo Rista Bleach Ombre	L'Oreal	NA			
Software and Algorithms					
Windows OBS Studio	Open Broadcast Software	https://obsproject.com/			
UMATracker	(Yamanaka & Takeuchi, 2018)	https://ymnk13.github.io/UMATracker/			
BORIS 8.0	Behavioral Observation Research Interactive Software	https://boris.readthedocs.io/en/latest/			
R v.3.6.3	The R Project for Statistical Computing	https://www.r-project.org/			
R package <i>trajr</i>	(McLean & Skowron Volponi, 2018)	https://cran.r- project.org/web/packages/trajr/index.html			
R package Ime4	(Bates, 2018)	https://cran.r-project.org/package=Ime4			
R package ImerTest	(Kuznetsova et al., 2017)	https://CRAN.R- project.org/package=ImerTest			
Adobe Photoshop	Adobe	https://www.adobe.com/			
ImageJ (Fiji)	NIH	https://imagej.net/software/fiji/			

Animals 404

All experimental subjects in this study were males (n=62) from two wild-derived partially inbred strains (NY2 and 405 406 NY3) of house mice (Mus musculus domesticus). Parental generations of these strains were caught in Saratoga Springs, NY by M. Sheehan (Phifer-Rixey et al., 2018). Wild-derived strains were used because competitive 407 behaviors characteristic of wild mice are less pronounced in highly inbred and domesticated laboratory strains 408 409 (Chalfin et al., 2014: Tuttle et al., 2018) and inbred strains tend to share identical urinary protein profiles (Cheetham et al., 2009). At weaning age (3-4 weeks) males were placed into a holding cage alone for 1-2 weeks, and were 410 411 subsequently paired with a female to allow for sexual experience as sexually naïve mice are known to exhibit 412 different social behavior (Stowers & Liberles, 2016). All males were allowed to reach adulthood, were between 3-5 413 months old by the time of experimental testing, and had the opportunity to produce one or more litters. All holding and breeding cages contained corn cob bedding, cardboard huts, and cotton nestlets. Mice were maintained in an 414 415 Animal Care facility at Cornell University with a 14:10 shifted light:dark cycle (lights went out at 10 PM and on at 12 416 PM) and were provided food and water ad libitum. Mice were handled minimally and with transfer cups whenever 417 possible to reduce stressful handling.

418 419 Urine collection

420 Urine was collected from each male subject and from C57BL/6 males to present self, familiar male (paired competitor), and unfamiliar male (C57BL/6) urine in the urine-marked zones on the final day of the trial series 421 (Figure 1A). Urine collection was performed using the single animal method: males were placed atop a metal grate 422 (an upside down cage hopper) over a clear plastic bag for 30 minutes to 1 hour (Kurien & Scofield, 1999). Males 423 424 were subsequently taken off the plastic bag and returned to their breeding cage. The urine droplets present on the 425 plastic bag were collected and stored at -80°C until use. Urine collected from subject males was stored individually 426 until the day of the urine-marked trials (Day 4: Figure 1A). For sufficient urine volume for the urine-marked trial treatments (Figure 1A), between 200-400uL was collected from each NY2 and NY3 male subject. On the day of the 427 428 urine-marked trials, individual aliquots for a subject male were thawed on ice and pooled together. Urine collection 429 for the C57BL/6 males was performed the month prior to beginning experimentation. We collected at least 50uL of 430 urine each from over 20 adult breeding C57BL/6 males, urine was stored on the day of collection at -80°C. Once a 431 sufficient volume was collected, individual aliguots were thawed on ice, and all C57BL/6 male urine was pooled into a single volume and subsequently aliquoted and stored at -80°C. This was done such that all unfamiliar C57BL/6 432 433 urine stimuli presented to males across trials were as similar as possible.

434

435 **Behavioral experiments**

All handling was performed with transfer cups throughout the duration of the trials to minimize stressful handling 436

- 437 confounds. One day prior to experimentation, we recorded subject male body weights to size-match individuals as
- closely as possible (average weight difference: 2.4g). All males were in breeding cages at the time of the 438
- experiment and most successfully reproduced (84%) prior to the trial series. As house mice are nocturnal, all 439 440 experiments were conducted in the dark during the dark cycle to ensure ethological accuracy (Peirson et al., 2018).

experimentation concluded prior to the winter months. Laboratory mice exhibit seasonal variation with respect to certain physiological parameters like serum concentrations of sex hormones, suggesting a possible mechanism for the internalization of annual time independent of light cycle, temperature, and humidity (Mock et al., 1975). While the available literature provides conflicting evidence as to whether these effects extend to behavior, we nonetheless took measures to avoid such confounds (Ferguson & Maier, 2013). Trial series were performed in sets of between 2-5 male pairs.

448 Behavioral trials consisted of a 4-day trial design, in which age and weight-matched adult breeding males of 449 distinct wild-derived strains (NY2 and NY3) were paired as competitors and presented a series of social and scent-450 marked trials (Figure 1A). We pair-matched each NY2 mouse with a NY3 mouse to ensure that no two paired mice 451 were genotypically identical and that their scent marks were perceptibly different (unique major urinary protein 452 profiles) (Kurien & Scofield, 1999), resulting in a total of 31 pairs (n=62). To ensure identification of males within a 453 pair (NY2 and NY3 strains are visibly indistinguishable), we ear-clipped and bleached a patch of rump fur of one 454 male in each pair a week prior to experimentation. Mice were anesthetized with isoflurane (5%). A heating pad was 455 used to maintain a stable body temperature. Isoflurane was delivered at 1-3% throughout the bleaching procedure. 456 L'Oreal Colo Rista Bleach Ombre (salon bleach) was mixed as per the manufacturer's instructions and dabbed onto 457 the top layer of fur using a sterile cotton swab. Care was taken to prevent bleach from contacting the skin. Twenty 458 minutes after application, sterile cotton tipped swabs dipped in water were used to rinse the bleach from the fur. 459 The fur was then dabbed dry with paper towels. Mice were placed under a heat lamp for 5 minutes or until they 460 were fully recovered from anesthesia before being transferred back to their home cage.

461 All trials were performed in one of two trial chambers that were sound proofed and fitted with recording 462 systems. For all trials large sheets of Whatman filter paper lined the floor of each trial to collect urine blots and to 463 present urine stimuli. The same size PVC arenas were used throughout (50 cm x 50 cm), though split in half with 464 the mesh barrier for the Mesh trials (Figure 1A). At the end of each trial, males were placed back into their breeding 465 cages. On Day 1 of the trial series, paired males were placed on either side of a wire mesh barrier in an arena for 466 30 minutes (Mesh 1, Figure 1A). At the end of the 30 minutes, males were briefly removed from the arena into large 467 transfer cups, the filter paper was labeled and removed, a fresh filter paper was placed in the arena, and the mesh 468 barrier was removed. Males were placed back into the arena for a 30 minute aggressive contest (Fight trial, Figure 469 1A). On Day 2, each male was placed alone in an empty "Open Field" arena without any stimuli aside from the 470 arena itself for 30 minutes (Open Field trial, Figure 1A). On Day 3, males were placed back into the Mesh arena for 471 30 minutes with the same male competitor encountered on the first day, without the subsequent fight trial (Mesh 2 472 trial, Figure 1A). On Day 4, males were placed into the arena alone and subjected to a 30 minute urine-marked 473 stimulus trial, consisting of one of 4 possible treatment types. Each treatment included two spatially distinct urine-474 marked zones placed in opposite corners of the arena (front right - back left vs. back right - front left). Urine-475 marked corner zones contained aliguoted male urine of 3 possible identities: self, familiar, or unfamiliar male. Self-476 urine was collected from the focal trial mouse; familiar male urine was from the paired male competitor of the focal 477 mouse; unfamiliar male urine was from pooled C57BL/6 urine. The urine stimuli for a urine-marked trial was 478 thawed, pooled and kept on ice until aliguoted for the urine-marked stimulus trial onto filter paper. Urine stimuli 479 were placed on the filter paper directly before the trial start in standardized locations and volumes. The four 480 treatment types span a range of scent mark combinations: self-self, self-familiar, self-unfamiliar, familiar-unfamiliar. 481 Paired males (winner-loser pairs) received the same urine-marked stimulus treatment. For all scent marked trials 482 (Days 1-4) the first and last minute of each trial was trimmed prior to analysis. This was done to minimize detection 483 of startle-based urination events caused by placement of mice into arenas and any jostling caused during trial set-484 up and take-down. The total analyzed trial length was thus 28 minutes.

485 Trials and treatments were randomized as follows. Male trial order and arena chamber was pseudo-486 randomized each day to avoid confounds in arena location and marking behavior over the course of the designated 487 trial period (12 PM – 4 PM). The orientation within the Mesh 1 trials was also randomized (whether males were 488 placed near the back or front of the arena) to account for variation in sound disturbances for males closer to the 489 chamber door; orientations were subsequently flipped for each pair in Mesh 2. Urine-marked trial treatments were 490 pseudo-randomly assigned to each male pair, to ensure similar numbers of male pairs were exposed to the 4 491 treatment types across sets of trials series. The orientation of urine stimuli was randomly assigned to corner 492 orientations (front right - back left vs. back right - front left). Lastly, the fur bleaching for male identification was 493 performed on one mouse strain (NY2 or NY3) for each trial set, but the bleached strain was switched between trial 494 sets to prevent errors within a trial set and to avoid bleaching only one strain across trial sets.

496 **Recording methods**

495

All trials were recorded with a security camera system (iDVR-PRO CMS) at 1080p and 30 frames per second to
visualize the high-speed aggressive encounters and to clearly distinguish the male identities (ear-marked and
bleached fur). All trials (including fight trials) were recorded thermally using an infrared camera system (PI 640;
Optris Infrared Sensing). Thermal cameras were fitted with 33° x 25° lenses and mounted above the experimental

arena chambers such that field-of-view for each camera covered the entire arena. The thermal detection window was set at: 61°F - 107°F. Data frames were collected at the max speed, averaging at 3 Hz. Thermal video data was saved by screen-capturing live Optris video output using OBS Studio software. Raw temperature data was also collected in semicolon-delimited CSVs, providing a readout of the temperature in each pixel for each frame.

506 Behavioral scoring

507 All videos were scored using Behavioral Observation Research Interactive Software (BORIS) (Friard & Gamba, 508 2016). For the fight trials (Figure 1A), we scored the following aggressive behaviors: chasing, hitting, boxing, and 509 wrestling bouts (Figure S1C) using the infrared security camera video recordings. To score urine mark deposition 510 events Optris thermal video recordings were used for all trials. Urine depositions were scored as a clear hot spot 511 following the focal mouse's trajectory that subsequently cooled below substrate temperature. Fecal depositions 512 could be eliminated as they are frequently cooler upon deposition event, cool much more slowly, have a distinct 513 shape, and are typically moved around the arena quickly. In addition to scoring the timing of urine deposition 514 events, the placement of urine marks was also scored. Using screen annotation software, we drew precise lines on 515 the video observation corresponding to regions of interest for each thermally-recorded trial (Figure 1B-C). 516

517 Tracking

505

518 Mice were tracked using the software UMATracker (Release 12) (Yamanaka & Takeuchi, 2018). Infrared security 519 camera recordings were used to track focal mouse movement, as the video were recorded at a higher framerate. 520 Filters were generated using the following modular settings (in order): output – Closing: Kernel = 6 – Opening: 521 Kernel = 6 – Threshold: 100 – BGRToGray – input. Videos were tracked using Group Tracker GMM algorithm. 522 Area51 was used to generate desired regions of interest for each trial (Figure 1B-C) and analyze the relative space 523 use in each of these regions. The R package *trajr* (McLean & Skowron Volponi, 2018) was used to quantitatively 524 characterize the following information from the tracked data frames: speed, acceleration, and trajectory length. 525

526 Urine blot imaging and processing

527 Trials were run on Whatman filter paper substrate. Arena edges were outlined with pencil on the filter paper at the 528 end of each trial. We collected all sheets of filter paper used in experimentation (except for the Fight trial) and 529 photographed them under ultraviolet (UV) light. We used three UV bulbs to evenly distribute light on the large filter 530 paper area. Images were converted to greyscale in Adobe Photoshop and the magentas were reduced to ~20% to 531 observe edges of urine marks clearly. Greyscale images were subsequently processed in ImageJ (Fiji). We 532 subtracted background pixels for a cleaner image (100 px), applied image thresholding (manually adjusted when 533 necessary), and converted images to binary in order to convert to mask, fill holes and perform watershed algorithm. 534 This processed image was then used to analyze the number of particles, with Size (pixel^2): 100-Infinity and 535 Circularity (0-1.00). 536

537 Urine mark bout classification

The median inter-mark interval (2.99 seconds) for all males across all trials was used to determine whether marks get clustered into a marking "bout" (Figure S4A). Any two marks that occur in sequence with an inter-mark interval less than 3 seconds are clustered together into a multi-mark bout, allowing us to examine within-bout temporal dynamics.

543 Statistical analysis

544 We conducted all statistical analyses in R 3.6.0 (R Development Core Team 2019). We used linear mixed models 545 (LMMs) and paired statistical tests to examine relationships between dependent and response variables. Models 546 were fitted using the package Ime4 (Bates, 2018). The ImerTest package was used to calculate degrees of freedom (Satterthwaite's method) and p-values (Kuznetsova et al., 2017). Dependent variables were transformed for a 547 548 subset of models to meet assumptions for model residuals after visually inspecting model residuals. We used a 549 type 3 analysis of variance (ANOVA) to test for overall effects of fixed factors or interactions in the models. Post 550 hoc comparisons were conducted using the emmeans package (Lenth, 2016). R script and data sheets used for all 551 statistical analyses are provided.

552

553 Acknowledgements

554 We thank Kevin Besler, Christen Rivera-Erick and Melanie Colvin for crucial technical assistance; Russell Ligon 555 and Caleb Vogt for helping establish recording systems and tracking methods in the lab; and James Tumulty for 556 manuscript feedback.

- 557
- 558

Funding				
	Funder	Grant reference number	Author	
	USDA Hatch Grant	NYC-191428	Michael J Sheehan	

The funders were not involved in the design of the study; the collection, analysis and interpretation of data; the writing of the manuscript and any decision concerning the publication of the paper.
 564

565 Author ORCIDs

566 Caitlin H Miller <u>https://orcid.org/0000-0002-3317-1821</u>

Additional information

567 Michael J Sheehan https://orcid.org/0000-0002-3949-7873

568 Melissa R Warden https://orcid.org/0000-0003-2240-3997

570 Ethics

571 All experimental protocols conducted at Cornell University were approved by the Institutional Animal Care and Use 572 Committee (IACUC: Protocol #2015-0060) and were in compliance with the NIH Guide for Care and Use of 573 Animals.

575 Authors' contributions.

576 CHM and MJS conceived the study. CHM performed trials and analyses. MFH, JY, BCC, KH and AYL collected 577 samples, scored behavioral trials, and generated tracking data. CHM and MJS wrote the initial drafts of the paper. 578 MRW edited the manuscript. All authors contributed to manuscript preparation.

Competing interests

581 The authors declare no competing interests.

583 Additional files

585 Supplementary figures



Figure S1. Male aggressive behaviors scored in contests (fight trials) between paired competitors. (A) Total aggressive behaviors performed by each paired male competitor. The fight outcome (the categorization of winners and losers) was determined by which male performed more aggressive behaviors within a pair. (B) Across all 31 pairs, winning males performed significantly more aggressive behaviors than losing males ($t_{1,37}$ = -12.6, p = 1.09e-13). Welch's t-test was used to compare the total aggressive behaviors performed by the two fight outcome categories (significance code: *** p<0.001). (C) Ethogram used to score aggressive behaviors. State events: chase, boxing and wrestling bouts. Points events: hits. An event is only coded for a male subject if the individual initiated the behavior (i.e. wrestling bout is coded for only one participant – the initiator – of that event).



Figure S2. Comparison of urine mark detection methods across trial types: Ultraviolet light (UV) blot imaging vs. thermal imaging. The two detection methods are well-correlated with each other (R > 0.8). For both Mesh trials and the Open Field trials, UV imaging consistently detected more urine marks than thermal imaging. The Marked trials revealed the opposite pattern, with thermal imagining detecting more urine marks than UV imaging. Three trials were excluded from this dataset due to poor urine blot quality, and one trial was excluded as an outlier.



Figure S3. Mesh trial spatial marking and space use. **(A)** Top: schematic of the mesh trials indicating the social "Barrier" (yellow) and non-social "Wall" regions of interest (ROIs). Below: Example urine blots of a male pair (winner and loser) pre- and post-fight demonstrating the spatial allocation of urine marks at the social boundary. **(B)** Linear mixed model (LMM) prediction of the total number of marks in the post-fight mesh trial (Mesh 2) given the fight outcome (winner=red; loser=blue), initial signal investment (# Mesh 1 marks), and the ROI (barrier=solid; wall=dashed). **(C)** Difference in time (s) spent in the Barrier vs. Wall regions of interest (ROIs) across mesh trials by winning and losing males. Winners and losers spend more time at the social boundary (Barrier) across mesh trials. Top left corner: an example heat map a male pair in a mesh trial (Mesh 1), depicting how all males spend more time at the social boundary (Barrier) than the non-social ROI (Wall) across mesh trials, regardless of fight outcome. **(D)** Comparison of the difference in time spent vs. the difference in total marks allocated in the ROIs (Barrier – Wall) by winners and loser across trials. In both mesh trials, space use and changes in urine allocation effort are not detectably correlated with each other among winning or losing males (R < 0.2). **(B-D)** Linear mixed models (LMMs) were used to model relationships, analyses of variance (ANOVAs) were used to test for overall effects, and post hoc pairwise comparisons were performed using the *emmeans* package (significance codes: NS p>0.05, * p<0.05; ** p<0.01, *** p<0.001).

bioRxiv preprint doi: https://doi.org/10.1101/2022.01.28.478242; this version posted January 28, 2022. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

17



Figure S4. (A) Histogram of the inter-mark intervals (IMIs) for all males across all trials. The median value is indicated with a dashed line (2.99 seconds). The range of IMIs extends to nearly the full trial length (only the first 50s is shown), The maximum values are reported in the bottom right corner. The median IMI value was used to define a marking "bout." Such that any two marks that occur in sequence with an IMI < 3 seconds are grouped together into a multi-mark bout. **(B)** Event plots depicting the urine marking of all male competitors over the course of both mesh trials (Mesh 1=left, Mesh 2 = right) for the entire trial duration (1800 seconds). Pair IDs are indicated on the left-hand axis. Losers depicted on top in blue, and winners on the bottom in red. **(C)** Event plots depicted a zoomed-in view of the first 200 seconds of the trials for all individuals. Example chain-like bouts are outlined in the Mesh 1 panel, and example burst-like marking bouts are highlighted in the Mesh 2 panel (yellow boxes).

bioRxiv preprint doi: https://doi.org/10.1101/2022.01.28.478242; this version posted January 28, 2022. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.



Figure S5. (A) Total number of marks deposited by winners and losers in scent-marked trials to a specific ROIs: scent-marked corners (containing self, familiar, or unfamiliar male urine), empty corners, or the center of the arena (significance codes: * p<0.05; ** p<0.01). (B) The percent of time spent in by winners and losers in specific urine-marked trial ROIs, normalized to the total area of each ROI (to account for the center being a larger area). Winner and losers spend significantly less time in the Center ROI than all corner ROIs (Self, Familiar, Unfamiliar or Empty; *** p<0.001). Losers spend significantly less time in the Center ROI than all corner ROIs (Self, Familiar, Unfamiliar corners relative to Empty ones (* p<0.05). (A,B) Linear mixed models (LMMs) were used to model relationships, analyses of variance (ANOVAs) were used to test for overall effects, and post hoc pairwise comparisons were performed using the *emmeans* package. (C) Event plots depicting the urine marking of winning and losing males to the OFTs and the urine-marked trials for the entire trial duration (1800 seconds). Males are grouped by the four different scent-marked treatments: self-self (S-S: light purple), self-familiar male (S-FM: dark purple), self-unfamiliar male (S-UM: light orange), and familiar male (TM-UM: dark orange). Almost all male pairs experienced the same treatment, three pairs received different urine-marked treatments due to urine stimuli collection constraints (hence some of the treatment groups have unequal paired number across fight outcome groupings).

References

- Alberts, A. C. (1992). Constraints on the Design of Chemical Communication Systems in Terrestrial Vertebrates. *The American Naturalist*, 139, S62–S89. https://doi.org/10.1086/285305
- Anderson, P. K., & Hill, J. L. (1965). Mus musculus: Experimental Induction of Territory Formation. *Science*, *148*(3678), 1753–1755. https://doi.org/10.1126/science.148.3678.1753
- Arakawa, H., Arakawa, K., Blanchard, D. C., & Blanchard, R. J. (2008). A new test paradigm for social recognition evidenced by urinary scent marking behavior in C57BL/6J mice. *Behavioural Brain Research*, *190*(1), 97–104. https://doi.org/10.1016/j.bbr.2008.02.009
- Arakawa, H., Blanchard, D. C., Arakawa, K., Dunlap, C., & Blanchard, R. J. (2008). Scent marking behavior as an odorant communication in mice. *Neuroscience & Biobehavioral Reviews*, *32*(7), 1236–1248. https://doi.org/10.1016/j.neubiorev.2008.05.012
- Arnott, G., & Elwood, R. W. (2009). Assessment of fighting ability in animal contests. *Animal Behaviour*, 77(5), 991–1004. https://doi.org/10.1016/j.anbehav.2009.02.010
- Aubin Horth, N., & Dodson, J. J. (2004). Influence of individual body siz and variable thresholds on the incidence of a sneaker male reproductive tactic in Atlantic salmon. *Evolution*, *58*(1), 136–144. https://doi.org/10.1111/j.0014-3820.2004.tb01580.x
- Bates, D. (2018). Parsimonious Mixed Models. ArXiv, arXiv:1506.04967, 21.
- Bhandiwad, A. A., Whitchurch, E. A., Colleye, O., Zeddies, D. G., & Sisneros, J. A. (2017). Seasonal plasticity of auditory saccular sensitivity in "sneaker" type II male plainfin midshipman fish, Porichthys notatus. *Journal of Comparative Physiology A*, 203(3), 211–222. https://doi.org/10.1007/s00359-017-1157-9
- Bittner, N. K. J., Mack, K. L., & Nachman, M. W. (2021). Gene expression plasticity and desert adaptation in house mice*. *Evolution*, 75(6), 1477–1491. https://doi.org/10.1111/evo.14172
- Booksmythe, I., Jennions, M. D., & Backwell, P. R. Y. (2010). Investigating the 'dear enemy' phenomenon in the territory defence of the fiddler crab, Uca mjoebergi. *Animal Behaviour*, *79*(2), 419–423. https://doi.org/10.1016/j.anbehav.2009.11.020
- Briefer, E., Rybak, F., & Aubin, T. (2008). When to be a dear enemy: Flexible acoustic relationships of neighbouring skylarks, Alauda arvensis. *Animal Behaviour*, 76(4), 1319–1325. https://doi.org/10.1016/j.anbehav.2008.06.017
- Briffa, M., & Elwood, R. W. (2009). Difficulties remain in distinguishing between mutual and self-assessment in animal contests. *Animal Behaviour*, 77(3), 759–762. https://doi.org/10.1016/j.anbehav.2008.11.010
- Chalfin, L., Dayan, M., Levy, D. R., Austad, S. N., Miller, R. A., Iraqi, F. A., Dulac, C., & Kimchi, T. (2014). Mapping ecologically relevant social behaviours by gene knockout in wild mice. *Nature Communications*, *5*(1), 4569. https://doi.org/10.1038/ncomms5569
- Cheetham, S. A., Smith, A. L., Armstrong, S. D., Beynon, R. J., & Hurst, J. L. (2009). Limited variation in the major urinary proteins of laboratory mice. *Physiology & Behavior*, *96*(2), 253–261. https://doi.org/10.1016/j.physbeh.2008.10.005
- Cheetham, S. A., Thom, M. D., Jury, F., Ollier, W. E. R., Beynon, R. J., & Hurst, J. L. (2007). The Genetic Basis of Individual-Recognition Signals in the Mouse. *Current Biology*, *17*(20), 1771–1777. https://doi.org/10.1016/j.cub.2007.10.007
- Christensen, C., & Radford, A. N. (2018). Dear enemies or nasty neighbors? Causes and consequences of variation in the responses of group-living species to territorial intrusions. *Behavioral Ecology*, 29(5), 1004–1013. https://doi.org/10.1093/beheco/ary010
- Cooper, B. G., & Goller, F. (2004). Multimodal Signals: Enhancement and Constraint of Song Motor Patterns by Visual Display. *Science*, 303(5657), 544–546. https://doi.org/10.1126/science.1091099
- Crowcroft, P., & Rowe, F. P. (1963). SOCIAL ORGANIZATION AND TERRITORIAL BEHAVIOUR IN THE. Proceedings of the Zoological Society of London, 140(3), 517–531. https://doi.org/10.1111/j.1469-7998.1963.tb01871.x
- Desjardins, C., Maruniak, J. A., & Bronson, F. H. (1973). Social Rank in House Mice: Differentiation Revealed by Ultraviolet Visualization of Urinary Marking Patterns. *Science*, *182*(4115), 939–941. https://doi.org/10.1126/science.182.4115.939
- Drickamer, L. C. (2001). Urine marking and social dominance in male house mice (Mus musculus domesticus). Behavioural Processes, 53(1–2), 113–120. https://doi.org/10.1016/S0376-6357(00)00152-2
- Dugatkin, L. A. (1997). Winner and loser effects and the structure of dominance hierarchies. *Behavioral Ecology*, 8(6), 583–587. https://doi.org/10.1093/beheco/8.6.583
- Enquist, M., & Leimar, O. (1983). Evolution of fighting behaviour: Decision rules and assessment of relative strength. Journal of Theoretical Biology, 102(3), 387–410. https://doi.org/10.1016/0022-5193(83)90376-4
- Ferguson, S. A., & Maier, K. L. (2013). A review of seasonal/circannual effects of laboratory rodent behavior. *Physiology* & *Behavior*, *119*, 130–136. https://doi.org/10.1016/j.physbeh.2013.06.007
- Ferkin, M. H. (2015). The response of rodents to scent marks: Four broad hypotheses. *Hormones and Behavior*, 68, 43– 52. https://doi.org/10.1016/j.yhbeh.2014.10.002
- Ferkin, M. H. (2019). Scent marks of rodents can provide information to conspecifics. *Animal Cognition*, 22(3), 445–452. https://doi.org/10.1007/s10071-019-01250-9

- 20
- Fertig, D. S., & Edmonds, V. W. (1969). The Physiology of the House Mouse. *Scientific American*, 221(4), 103–110. https://doi.org/10.1038/scientificamerican1069-103
- Friard, O., & Gamba, M. (2016). BORIS: A free, versatile open-source event-logging software for video/audio coding and live observations. *Methods in Ecology and Evolution*, 7(11), 1325–1330. https://doi.org/10.1111/2041-210X.12584
- Gil, D., & Gahr, M. (2002). The honesty of bird song: Multiple constraints for multiple traits. *Trends in Ecology & Evolution*, 17(3), 133–141. https://doi.org/10.1016/S0169-5347(02)02410-2
- Goll, Y., Demartsev, V., Koren, L., & Geffen, E. (2017). Male hyraxes increase countersinging as strangers become 'nasty neighbours.' *Animal Behaviour*, 134, 9–14. https://doi.org/10.1016/j.anbehav.2017.10.002
- Gosling, L. M. (1982). A Reassessment of the Function of Scent Marking in Territories. *Zeitschrift Für Tierpsychologie*, 60(2), 89–118. https://doi.org/10.1111/j.1439-0310.1982.tb00492.x
- Gosling, L. M., Roberts, S. C., Thornton, E. A., & Andrew, M. J. (2000). Life history costs of olfactory status signalling in mice. *Behavioral Ecology and Sociobiology*, 48(4), 328–332. https://doi.org/10.1007/s002650000242
- Harrington, J. E. (1976). Recognition of Territorial Boundaries by Olfactory Cues in Mice (Mus musculus L.). Zeitschrift *Für Tierpsychologie*, 41(3), 295–306. https://doi.org/10.1111/j.1439-0310.1976.tb00484.x
- Harrison, L. M., Jennions, M. D., & Head, M. L. (2018). Does the winner–loser effect determine male mating success? Biology Letters, 14(5), 20180195. https://doi.org/10.1098/rsbl.2018.0195
- Hobson, E. A. (2020). Differences in social information are critical to understanding aggressive behavior in animal dominance hierarchies. *Current Opinion in Psychology*, 33, 209–215. https://doi.org/10.1016/j.copsyc.2019.09.010
- Hou, X. H., Hyun, M., Taranda, J., Huang, K. W., Todd, E., Feng, D., Atwater, E., Croney, D., Zeidel, M. L., Osten, P., & Sabatini, B. L. (2016). Central Control Circuit for Context-Dependent Micturition. *Cell*, 167(1), 73-86.e12. https://doi.org/10.1016/j.cell.2016.08.073
- Hsu, Y., & Wolf, L. L. (1999). The winner and loser effect: Integrating multiple experiences. *Animal Behaviour*, 57(4), 903–910. https://doi.org/10.1006/anbe.1998.1049
- Humphries, E. L., Hebblethwaite, A. J., Batchelor, T. P., & Hardy, I. C. W. (2006). The importance of valuing resources: Host weight and contender age as determinants of parasitoid wasp contest outcomes. *Animal Behaviour*, 72(4), 891–898. https://doi.org/10.1016/j.anbehav.2006.02.015
- Hurst, J. L. (1990). Urine marking in populations of wild house mice Mus domesticus rutty. I. Communication between males. *Animal Behaviour*, 40(2), 209–222. https://doi.org/10.1016/S0003-3472(05)80916-9
- Hurst, J. L., & Beynon, R. J. (2004). Scent wars: The chemobiology of competitive signalling in mice. *BioEssays*, 26(12), 1288–1298. https://doi.org/10.1002/bies.20147
- Hurst, J. L., Payne, C. E., Nevison, C. M., Marie, A. D., Humphries, R. E., Robertson, D. H. L., Cavaggioni, A., & Beynon, R. J. (2001). Individual recognition in mice mediated by major urinary proteins. *Nature*, 414(6864), 631–634. https://doi.org/10.1038/414631a
- Hurst, J. L., Thom, M. D., Nevison, C. M., Humphries, R. E., & Beynon, R. J. (2005). MHC odours are not required or sufficient for recognition of individual scent owners. *Proceedings of the Royal Society B: Biological Sciences*, 272(1564), 715–724. https://doi.org/10.1098/rspb.2004.3004
- Jin, L., Liang, J., Fan, Q., Yu, J., Sun, K., & Wang, H. (2021). Male Great Tits (Parus major) adjust dear enemy effect expression in different breeding stages. *Journal of Ornithology*, *162*(1), 221–229. https://doi.org/10.1007/s10336-020-01815-3
- Johnstone, R. (1996). Multiple displays in animal communication: 'Backup signals' and 'multiple messages.' *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 351(1337), 329–338. https://doi.org/10.1098/rstb.1996.0026
- Jones, R. B., & Nowell, N. W. (1973). Aversive and aggression-promoting properties of urine from dominant and subordinate male mice. *Animal Learning & Behavior*, *1*(3), 207–210. https://doi.org/10.3758/BF03199075
- Kaur, A. W., Ackels, T., Kuo, T.-H., Cichy, A., Dey, S., Hays, C., Kateri, M., Logan, D. W., Marton, T. F., Spehr, M., & Stowers, L. (2014). Murine Pheromone Proteins Constitute a Context-Dependent Combinatorial Code Governing Multiple Social Behaviors. *Cell*, 157(3), 676–688. https://doi.org/10.1016/j.cell.2014.02.025
- Keller, J. A., Chen, J., Simpson, S., Wang, E. H.-J., Lilascharoen, V., George, O., Lim, B. K., & Stowers, L. (2018). Voluntary urination control by brainstem neurons that relax the urethral sphincter. *Nature Neuroscience*, 21(9), 1229–1238. https://doi.org/10.1038/s41593-018-0204-3
- Kodric-Brown, A., & Brown, J. H. (1984). Truth in Advertising: The Kinds of Traits Favored by Sexual Selection. *The American Naturalist*, 124(3), 309–323. https://doi.org/10.1086/284275
- Koolhaas, J. M., Coppens, C. M., de Boer, S. F., Buwalda, B., Meerlo, P., & Timmermans, P. J. A. (2013). The Residentintruder Paradigm: A Standardized Test for Aggression, Violence and Social Stress. *Journal of Visualized Experiments*, 77, 4367. https://doi.org/10.3791/4367
- Kurien, B. T., & Scofield, R. H. (1999). Mouse urine collection using clear plastic wrap. *Laboratory Animals*, 33(1), 83–86. https://doi.org/10.1258/002367799780578525
- Kuznetsova, A., Brockhoff, P. B., & Christensen, R. H. B. (2017). ImerTest Package: Tests in Linear Mixed Effects Models. *Journal of Statistical Software*, 82(13). https://doi.org/10.18637/jss.v082.i13
- Laidre, M. E., & Johnstone, R. A. (2013). Animal signals. *Current Biology*, *23*(18), R829–R833. https://doi.org/10.1016/j.cub.2013.07.070

- Lee, W., Khan, A., & Curley, J. P. (2017). Major urinary protein levels are associated with social status and context in mouse social hierarchies. *Proceedings of the Royal Society B: Biological Sciences*, 284(1863), 20171570. https://doi.org/10.1098/rspb.2017.1570
- Lenth, R. V. (2016). Least-Squares Means: The *R* Package **Ismeans**. *Journal of Statistical Software*, 69(1). https://doi.org/10.18637/jss.v069.i01
- Li, C.-Y., Earley, R. L., Huang, S.-P., & Hsu, Y. (2014). Fighting experience alters brain androgen receptor expression dependent on testosterone status. *Proceedings of the Royal Society B: Biological Sciences*, 281(1796), 20141532. https://doi.org/10.1098/rspb.2014.1532
- Ligon, R. A., & McGraw, K. J. (2016). Social costs enforce honesty of a dynamic signal of motivation. *Proceedings of the Royal Society B: Biological Sciences*, 283(1841), 20161873. https://doi.org/10.1098/rspb.2016.1873
- Mackintosh, J. H. (1970). *Territory formation by laboratory mice*. *18*, 177–183. https://doi.org/10.1016/0003-3472(70)90088-6
- McLean, D. J., & Skowron Volponi, M. A. (2018). trajr: An R package for characterisation of animal trajectories. *Ethology*, 124(6), 440–448. https://doi.org/10.1111/eth.12739
- Miles, D. B., Sinervo, B., Hazard, L. C., Svensson, E. I., & Costa, D. (2007). Relating endocrinology, physiology and behaviour using species with alternative mating strategies. *Functional Ecology*, 21(4), 653–665. https://doi.org/10.1111/j.1365-2435.2007.01304.x
- Milewski, T. M., Lee, W., Champagne, F. A., & Curley, J. P. (2022). Behavioural and physiological plasticity in social hierarchies. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 377(1845), 20200443. https://doi.org/10.1098/rstb.2020.0443
- Mock, E. J., Kamel, F., Wright, W. W., & Frankel, A. I. (1975). Seasonal rhythm in plasma testosterone and luteinising hormone of the male laboratory rat. *Nature*, *256*(5512), 61–63. https://doi.org/10.1038/256061a0
- Moro, D., & Bradshaw, S. D. (1999). Water and sodium balances and metabolic physiology of house mice (Mus domesticus) and short-tailed mice (Leggadina lakedownensis) under laboratory conditions. *Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology*, 169(8), 538–548. https://doi.org/10.1007/s003600050253
- Müller, C. A., & Manser, M. B. (2007). 'Nasty neighbours' rather than 'dear enemies' in a social carnivore. *Proceedings of the Royal Society B: Biological Sciences*, 274(1612), 959–965. https://doi.org/10.1098/rspb.2006.0222
- Nelson, A. C., Cunningham, C. B., Ruff, J. S., & Potts, W. K. (2015). Protein pheromone expression levels predict and respond to the formation of social dominance networks. *Journal of Evolutionary Biology*, 28(6), 1213–1224. https://doi.org/10.1111/jeb.12643
- Nevison, C. M., Barnard, C. J., Beynon, R. J., & Hurst, J. L. (2000). The consequences of inbreeding for recognizing competitors. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 267(1444), 687–694. https://doi.org/10.1098/rspb.2000.1057
- Olivier, B., Mos, J., & Slangen, J. L. (Eds.). (1991). Animal Models in Psychopharmacology. Birkhäuser Basel. https://doi.org/10.1007/978-3-0348-6419-0
- Pasch, B., Tokuda, I. T., & Riede, T. (2017). Grasshopper mice employ distinct vocal production mechanisms in different social contexts. *Proceedings of the Royal Society B: Biological Sciences*, 284(1859), 20171158. https://doi.org/10.1098/rspb.2017.1158
- Patricelli, G. L., & Hebets, E. A. (2016). New dimensions in animal communication: The case for complexity. *Current Opinion in Behavioral Sciences*, *12*, 80–89. https://doi.org/10.1016/j.cobeha.2016.09.011
- Peirson, S. N., Brown, L. A., Pothecary, C. A., Benson, L. A., & Fisk, A. S. (2018). Light and the laboratory mouse. Journal of Neuroscience Methods, 300, 26–36. https://doi.org/10.1016/j.jneumeth.2017.04.007
- Phifer-Rixey, M., Bi, K., Ferris, K. G., Sheehan, M. J., Lin, D., Mack, K. L., Keeble, S. M., Suzuki, T. A., Good, J. M., & Nachman, M. W. (2018). The genomic basis of environmental adaptation in house mice. *PLOS Genetics*, 14(9), e1007672. https://doi.org/10.1371/journal.pgen.1007672
- Poole, T. B., & Morgan, H. D. R. (1976). Social and territorial behaviour of laboratory mice (Mus musculus L.) in small complex areas. *Animal Behaviour*, 24(2), 476–480. https://doi.org/10.1016/S0003-3472(76)80056-5
- Rauber, R., & Manser, M. B. (2018). Experience of the signaller explains the use of social versus personal information in the context of sentinel behaviour in meerkats. *Scientific Reports*, 8(1), 11506. https://doi.org/10.1038/s41598-018-29678-y
- Rose, J., Cullen, D. A., Simpson, S. J., & Stevenson, P. A. (2017). Born to win or bred to lose: Aggressive and submissive behavioural profiles in crickets. *Animal Behaviour*, *123*, 441–450. https://doi.org/10.1016/j.anbehav.2016.11.021
- Ruppé, L., Clément, G., Herrel, A., Ballesta, L., Décamps, T., Kéver, L., & Parmentier, E. (2015). Environmental constraints drive the partitioning of the soundscape in fishes. *Proceedings of the National Academy of Sciences*, *112*(19), 6092–6097. https://doi.org/10.1073/pnas.1424667112
- Rychlik, L., & Zwolak, R. (2005). Behavioural mechanisms of conflict avoidance among shrews. *Acta Theriologica*, *50*(3), 289–308. https://doi.org/10.1007/BF03192627
- Sethi, S., Lin, H.-H., Shepherd, A. K., Volkan, P. C., Su, C.-Y., & Wang, J. W. (2019). Social Context Enhances Hormonal Modulation of Pheromone Detection in Drosophila. *Current Biology*, *29*(22), 3887-3898.e4. https://doi.org/10.1016/j.cub.2019.09.045

Sheehan, M. J., Campbell, P., & Miller, C. H. (2019). Evolutionary patterns of major urinary protein scent signals in house mice and relatives. *Molecular Ecology*, 28(15), 3587–3601. https://doi.org/10.1111/mec.15155

- Sheehan, M. J., Lee, V., Corbett-Detig, R., Bi, K., Beynon, R. J., Hurst, J. L., & Nachman, M. W. (2016). Selection on Coding and Regulatory Variation Maintains Individuality in Major Urinary Protein Scent Marks in Wild Mice. *PLOS Genetics*, 12(3), e1005891. https://doi.org/10.1371/journal.pgen.1005891
- Sinervo, B., & Lively, C. M. (1996). The rock–paper–scissors game and the evolution of alternative male strategies. *Nature*, 380(6571), 240–243. https://doi.org/10.1038/380240a0
- Siracusa, E. R., Boutin, S., Dantzer, B., Lane, J. E., Coltman, D. W., & McAdam, A. G. (2021). Familiar Neighbors, but Not Relatives, Enhance Fitness in a Territorial Mammal. *Current Biology*, *31*(2), 438-445.e3. https://doi.org/10.1016/j.cub.2020.10.072
- Stowers, L., & Liberles, S. D. (2016). State-dependent responses to sex pheromones in mouse. *Current Opinion in Neurobiology*, 38, 74–79. https://doi.org/10.1016/j.conb.2016.04.001
- Sullivan-Beckers, L., & Hebets, E. A. (2014). Tactical adjustment of signalling leads to increased mating success and survival. *Animal Behaviour*, 93, 111–117. https://doi.org/10.1016/j.anbehav.2014.04.021
- Számadó, S. (2017). When honesty and cheating pay off: The evolution of honest and dishonest equilibria in a conventional signalling game. *BMC Evolutionary Biology*, *17*(1), 270. https://doi.org/10.1186/s12862-017-1112-y
- Thomas, A. L., Davis, S. M., & Dierick, H. A. (2015). Of Fighting Flies, Mice, and Men: Are Some of the Molecular and Neuronal Mechanisms of Aggression Universal in the Animal Kingdom? *PLOS Genetics*, *11*(8), e1005416. https://doi.org/10.1371/journal.pgen.1005416
- Tibbetts, E. A., & Crocker, K. C. (2014). The challenge hypothesis across taxa: Social modulation of hormone titres in vertebrates and insects. *Animal Behaviour*, *92*, 281–290. https://doi.org/10.1016/j.anbehav.2014.02.015
- Tibbetts, E. A., & Izzo, A. (2010). Social Punishment of Dishonest Signalers Caused by Mismatch between Signal and Behavior. *Current Biology*, 20(18), 1637–1640. https://doi.org/10.1016/j.cub.2010.07.042
- Tumulty, J. P. (2018). Dear Enemy Effect. In J. Vonk & T. Shackelford (Eds.), *Encyclopedia of Animal Cognition and Behavior* (pp. 1–4). Springer International Publishing. https://doi.org/10.1007/978-3-319-47829-6_693-1
- Tumulty, J. P., & Bee, M. A. (2021). Ecological and social drivers of neighbor recognition and the dear enemy effect in a poison frog. *Behavioral Ecology*, 32(1), 138–150. https://doi.org/10.1093/beheco/araa113
- Tuttle, A. H., Philip, V. M., Chesler, E. J., & Mogil, J. S. (2018). Comparing phenotypic variation between inbred and outbred mice. *Nature Methods*, *15*(12), 994–996. https://doi.org/10.1038/s41592-018-0224-7
- Verstegen, A. M. J., Klymko, N., Zhu, L., Mathai, J. C., Kobayashi, R., Venner, A., Ross, R. A., VanderHorst, V. G., Arrigoni, E., Geerling, J. C., & Zeidel, M. L. (2019). Non-Crh Glutamatergic Neurons in Barrington's Nucleus Control Micturition via Glutamatergic Afferents from the Midbrain and Hypothalamus. *Current Biology*, 29(17), 2775-2789.e7. https://doi.org/10.1016/j.cub.2019.07.009
- Wolff, R. J. (1985). Mating behaviour and female choice: Their relation to social structure in wild caught House mice (*Mus musculus*) housed in a semi-natural environment. *Journal of Zoology*, 207(1), 43–51. https://doi.org/10.1111/j.1469-7998.1985.tb04914.x
- Yamanaka, O., & Takeuchi, R. (2018). UMATracker: An intuitive image-based tracking platform. *Journal of Experimental Biology*, jeb.182469. https://doi.org/10.1242/jeb.182469
- Zamudio, K. R., & Sinervo, B. (2000). Polygyny, mate-guarding, and posthumous fertilization as alternative male mating strategies. *Proceedings of the National Academy of Sciences*, 97(26), 14427–14432. https://doi.org/10.1073/pnas.011544998
- Zorzal, G., Camarota, F., Dias, M., Vidal, D. M., Lima, E., Fregonezi, A., & Campos, R. I. (2021). The dear enemy effect drives conspecific aggressiveness in an Azteca-Cecropia system. *Scientific Reports*, *11*(1), 6158. https://doi.org/10.1038/s41598-021-85070-3